



Impact of Urban Runoff on Benthic and Pelagic Fish Fauna in Ikpoba River: Heavy Metals and Pathology of Liver Tissues

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Abstract

This study investigates the impact of urban runoff on benthic and pelagic fish fauna in the Ikpoba River, focusing on the effects of heavy metals on liver tissues. The research was conducted in Benin City, Nigeria, where the river is central to local livelihoods but faces pollution threats due to proximity to waste disposal and industrial effluent sites. Water, sediment, and fish samples were collected from two sampling stations along the river. The concentrations of heavy metals (Cr, Co, Cd, Ni, Pb) in these samples were analyzed using an Atomic Absorption Spectrophotometer. Histopathological examinations of fish liver tissues were also conducted. Results revealed the presence of heavy metals in the water and sediment samples, with significant concentrations of chromium upstream. Heavy metals were also found in the liver of both benthic and pelagic fishes, indicating potential environmental contamination. Histopathology of the liver tissues showed signs of inflammation and steatosis, common responses to stress or injury, such as that caused by heavy metal exposure. These findings underscore the need for ongoing monitoring and management of the river's health.

1.0. Introduction

Predisposition Urban runoff, originating from impermeable urban surfaces, poses a significant environmental challenge due to its role in contaminating water bodies with various pollutants [1, 2]. This runoff carries a range of contaminants such as heavy metals, toxic chemicals, microplastics, and pathogens, contributing to water quality degradation and ecosystem disruption [3, 4]. Additionally, urbanization alters hydrological processes, leading to increased storm runoff peaks and reduced baseflows, impacting streamflow regimes over time [5]

Urban runoff is a significant source of pollutants that can degrade surface water bodies, such as the Ikpoba River. Studies have shown consistent declines in the richness of algal, invertebrate, and fish communities in urban streams due to the impact of urban runoff [6, 7].

Heavy metals such as lead, cadmium, arsenic, and mercury pose significant risks to both aquatic ecosystems and human health due to their accumulation in fish tissues. Fish, being bio-monitors of pollution, play a crucial role in reflecting the levels of heavy metals in aquatic environments [8]. The liver, a vital organ for heavy metal accumulation, is particularly susceptible to the bioaccumulation of these pollutants [9]. Studies have shown that heavy metal concentrations in fish

tissues can vary based on factors like species, climate, and developmental stage, highlighting the complexity of heavy metal accumulation in fish [10].

Urban runoff can have a multifaceted impact on benthic and pelagic fish species, influenced by factors such as water depth, temperature, and food sources. Metals present in runoff can accumulate in fish tissues, posing health risks and inducing oxidative stress [11]. Excessive nutrients like phosphorus and nitrogen from runoff can trigger algal blooms, diminishing oxygen levels and endangering fish [12]. Sedimentation caused by runoff can exacerbate the issue by depleting dissolved oxygen, further impacting fish populations negatively ([6].

The liver of fish plays a crucial role in safeguarding against oxidative stress through essential antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx) [13-15]. Heavy metal contamination in aquatic ecosystems poses a significant threat to fish health, particularly impacting the liver tissues. Studies have shown that exposure to heavy metals like zinc, lead, copper, and cadmium can lead to histopathological alterations in fish liver tissues, including vascular congestion, hepatopancreas degeneration, necrosis, and hypertrophy of hepatocytes [16-18]. The accumulation of these toxic heavy metals in fish liver tissues has been well-documented, highlighting the severity of heavy metal pollution in aquatic environments [19]. This research seeks to explore the impact of urban runoff on fish populations in the Ikpoba River, with a particular emphasis on the effects of pollutants, specifically heavy metals and antioxidant enzymes, on benthic and pelagic fish species. The study will involve assessing the concentrations of these substances in fish livers, investigating the implications of urban runoff on the fish populations, evaluating the levels of metals in fish livers as indicators of pollution, and examining the correlation between urban runoff and fish populations by determining the influence of harmful pollutants on the health of the fishes [20].

The Ikpoba River in Nigeria's Benin-Owena basin is a vital natural resource for the local community, supporting various activities [21, 22]. It serves as an important aquatic ecosystem, providing habitat for diverse fish species and maintaining water quality [23, 24]. Economically, the river sustains fishing activities, offering income and food for residents, and supports waste management and industrial activities [25-27]. Culturally, it is significant for various practices, and its health is valued by the community [25, 27]. However, concerns about pollution from waste and industrial sites threaten its importance, highlighting the need to protect its ecological, economic, and cultural value for sustainable community development [28, 29].

The scope of the research will be focused on the influence of urban runoff on benthic and pelagic fish populations in the Ikpoba River. This will entail the analysis of physicochemical characteristics, the collection of fish samples from both habitats, and the examination of liver samples to assess antioxidant enzymes, oxidative stress, and variations in heavy metal concentrations. The findings from this study will provide valuable insights into the repercussions of urban runoff on the fish fauna in the Ikpoba River, enhance our understanding of the associated health risks, and contribute to the development of strategies for protecting aquatic ecosystems from the adverse effects of urban runoff, identifying potential sources of contamination, and suggesting mitigation measures.

2.0. Materials and Methods

The study took place in Benin City, a southern Nigeria within Edo State, framed by latitudes 6°11' N to 6°29' N and longitudes 5°33' E to 5°47' E. The city's terrain, perched at roughly 78 meters above sea level, is part of Nigeria's rich and humid tropical rainforest region. The Ikpoba River, a significant natural resource, flows through the Benin-Owena basin, intersecting Egor and Ikpoba-Okha [30]. It is a vital artery for the community, supporting fishing, waste management, recreational activities, and industrial effluent disposal [31].

The river is central to the livelihoods of the local populace, facilitating fishing practices and bearing the brunt of human-induced pressures that threaten its water quality and ecosystem vitality health [30, 32]. The proximity of waste disposal and industrial effluent to the river's banks has sparked

concerns over pollution and ecological damage. The interplay between human actions and the river's ecological health calls for integrated environmental management approaches that ensure sustainable development while protecting the river's integrity.

2.1. Collection of samples

The study was conducted at two sampling stations along the Ikpoba River, with water, sediment, and fish samples collected for analysis.

2.1.1. Water Samples

Samples were gathered from both the benthic and pelagic zones of the Ikpoba River during regular expeditions from June to August 2023. The method described by Eric, et al. [33] was followed, using thoroughly cleaned 1-liter sampling containers.. After collection, the bottles were immediately sealed, marked, and kept on ice for transport to the lab for further investigation [31].

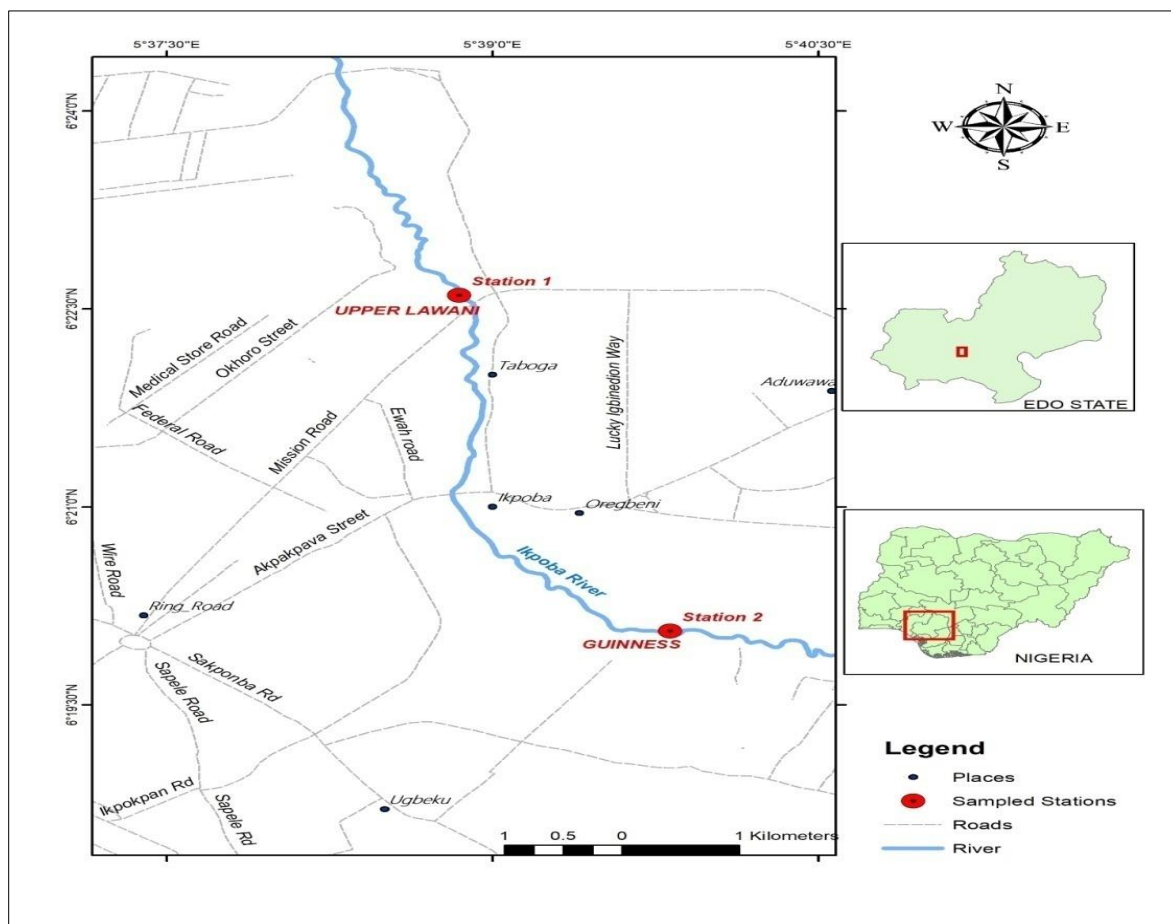


Figure 1: Map showing site locations of the study area

2.1.2. Sediment Sampling

The grab sampling technique was employed for sediment collection. A porcelain crucible was used to weigh ten grams (10 g) of ground and sieved sediment sample. Following this, 25 ml of distilled water, 1 ml of concentrated nitric acid (HNO_3), and 3 ml of concentrated hydrochloric acid (HCl) were added. The sample was then heated on a steam bath for approximately an hour and subsequently cooled. The digested sample was filtered and adjusted to 50 ml with distilled water.

The filtrate was stored in plastic containers for later heavy metal analysis, ensuring consistent treatment procedures for all sediment samples [31].

2.1.3. Fish Sampling: A fishing net was used to randomly collect fish specimens based on the method described by Tongo, et al. [34]. A total of fifty-four (54) fish were collected, with thirty from the pelagic zone and twenty-four from the benthic zone. These specimens were carefully transported to the laboratory for further analysis.

2.1.4. Laboratory Procedure In the controlled laboratory setting, we meticulously cleaned the fish using distilled water. To ensure proper identification and preservation, each fish was stored separately in polyethylene bags at a temperature of -10°C .

2.1.5. Heavy Metal Analysis

The presence of heavy metals in water and sediment samples was ascertained using the techniques outlined by Davies and Ekperusi [35] and Osioma and Iniaghe [36]. A 25ml water sample was placed into a porcelain crucible, followed by the addition of 1ml of concentrated nitric acid (HNO_3) and 3ml of concentrated hydrochloric acid (HCl). The mixture was heated on a steam bath for roughly 30 minutes and then cooled. The resulting digest was diluted to 50ml with distilled water and stored in plastic containers for subsequent heavy metal analysis, ensuring a uniform procedure for all water samples.

In the case of sediment analysis, a 10g sample of ground and sieved sediment was placed into a porcelain crucible. Then, 25ml of distilled water was added, along with 1ml of concentrated nitric acid (HNO_3) and 3ml of concentrated hydrochloric acid (HCl). The sample was heated on a steam bath for about an hour and then cooled. The digested sample was filtered and adjusted to 50ml with distilled water. The filtrate was stored in plastic containers for later heavy metal analysis, ensuring a consistent procedure for all sediment samples.

Fish samples were meticulously measured for length and weight, and the liver was surgically removed, weighed, and set aside for heavy metal determination. The samples were digested with 1ml of concentrated nitric acid (HNO_3) and 3ml of concentrated hydrochloric acid (HCl) until a clear and transparent solution was obtained based on the method described by Jabeen, et al. [37]. The digested samples were diluted to the required volume using double distilled water in a 500 ml volumetric flask. After wet digestion, fish organ samples were analyzed for heavy metals.

The heavy metals in the water, sediment, and fish samples were analyzed using an Atomic Absorption Spectrophotometer (AAS) Solar 969 Unicam Series model. Each metal (Cr, Co, Cd, Ni, Pb) was determined using a specific hollow cathode lamp for its analysis. Each sample was analyzed three times to ensure representative results, and the metal concentrations were calculated using a standard calibration plot method, as described by Chukwuka and Ogbeide [31].

2.1.6. Histopathological Examination of Fish Liver

Liver tissues were preserved in a 10% buffer solution for a day, then dissected into small pieces and positioned in a tissue cassette based on the method described by Anyiamuka-Chinedu, et al. [38]. The samples were subsequently washed and subjected to an Automatic Tissue Processing (ATP) machine for an hour, where they stayed in the processing solutions for half a day before being extracted from the ATP machine. The tissue samples were then embedded by situating them in paraffin wax blocks, which offered structural support. Sections of 3 - 5 microns were created through microtomy and made to float on water in a floatation bath at 56°C to eliminate wrinkles, and then relocated to microscopic slides. The tissues underwent a xylene treatment to eliminate the paraffin wax, followed by dehydration using ethanol. They were then stained with haematoxylin and eosin stains and finally scrutinized under a light microscope at a magnification of $\times 100$ [39].

2.2. Statistical Analysis

The data gathered from the analysis was processed statistically using the SPSS software, version 21. The analyzed data was displayed in summary tables as Mean \pm S.E. For the examination of measurements within each sampling location, a One-way ANOVA and Duncan Multiple Range (DMR) test will be employed.

3.0. Results

3.1. Concentration of Metals in Water from Ikpoba River

The table 1 present data on the monthly concentrations of heavy metals in water samples collected from the Ikpoba River. Over the months of June, July, and August, the concentrations of nickel, chromium, and cobalt showed variations between upstream and downstream locations, while the levels of lead and cadmium remained constant at 0 mg/L. The general concentration of heavy metals, as depicted in Table 1 highlights a significant presence of chromium upstream. The mean difference is significant at $P < 0.05$.

Table 1: Monthly concentrations of heavy metals in water

Heavy Metal	June		July		August	
	Upstream	Down stream	Upstream	Down stream	Upstream	Down stream
NICKEL	0.04 \pm 0.002	0.05 \pm 0.002	0.03 \pm 0.004	0.05 \pm 0.002	0.03 \pm 0.005	0.04 \pm 0.002
LEAD	0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0
CHROMIUM	0.03 \pm 0.003	0.05 \pm 0.003	0.04 \pm 0.001	0.04 \pm 0.003	0.04 \pm 0.004	0.05 \pm 0.003
CADMIUM	0.0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0	0.0 \pm 0.0
COBALT	0.02 \pm 0.003	0.02 \pm 0.006	0.02 \pm 0.003	0.03 \pm 0.002	0.03 \pm 0.004	0.04 \pm 0.002

The values are the average standard deviation in three independent analysis. All values are in mg/L. Mean difference is significant at $P < 0.05$

3.2. Concentration of Metals in Sediment from Ikpoba River

Table 2 shows monthly heavy metal concentrations in Ikpoba River sediments. Nickel (Ni) had the highest concentration (7.89 mg/g downstream in July), while Cadmium (Cd) had the lowest (0.177 mg/g upstream in August). Lead (Pb) increased significantly in August (5.827 mg/g upstream, 5.208 mg/g downstream), while Cobalt (Co) and Ni decreased. Chromium (Cr) and Cd concentrations remained stable. Differences in Pb, Co, and Ni concentrations were statistically significant ($P < 0.05$), unlike Cr and Cd.

Table 2: Monthly Concentrations(mg/g) of Heavy Metals in Sediments from Ikpoba River

Heavy metal	June		July		August		P value
	Upstream	Downstream	Upstream	Downstream	Upstream	Downstream	
	Mean \pm S.E	Mean \pm S.E	Mean \pm S.E	Mean \pm S.E	Mean \pm S.E	Mean \pm S.E	
Pb	0.362 \pm 0.019	0.474 \pm 0.004	0.362 \pm 0.019	0.474 \pm 0.004	5.827 \pm 0.595	5.208 \pm 0.348	$P < 0.05$
Co	1.718 \pm 0.136	2.067 \pm 0.058	1.718 \pm 0.136	2.067 \pm 0.058	0.489 \pm 0.019	0.513 \pm 0.045	$P < 0.05$
Cr	2.568 \pm 0.297	2.155 \pm 0.021	2.568 \pm 0.297	2.155 \pm 0.021	1.899 \pm 0.149	2.116 \pm 0.342	$P > 0.05$
Cd	0.181 \pm 0.004	0.214 \pm 0.002	0.181 \pm 0.004	0.214 \pm 0.002	0.177 \pm 0.036	0.241 \pm 0.022	$P > 0.05$
Ni	6.125 \pm 0.102	7.89 \pm 0.193	6.125 \pm 0.102	7.89 \pm 0.193	1.585 \pm 0.061	1.447 \pm 0.117	$P < 0.05$

The values are the average standard deviation in three independent analyses. All values are in mg/kg. Mean difference is significant at $P < 0.05$

3.3. Concentration of Metals in Fish Liver from Ikpoba River

The monthly concentrations of heavy metals in the liver of benthic fishes showed some fluctuations. In May and June, the concentrations of lead, cobalt, chromium, cadmium, and nickel were 0.0618 mg/g, 0.073 mg/g, 0.348 mg/g, 0.0494 mg/g, and 0.8102 mg/g respectively. However, in July, there was a significant increase in the concentration of lead to 0.864 mg/g, while the concentrations of cobalt, chromium, and nickel decreased to 0.06175 mg/g, 0.4885 mg/g, and 0.0665 mg/g respectively. The concentration of cadmium also decreased slightly to 0.0335 mg/g.

For pelagic fishes, the concentrations of heavy metals in the liver also varied. In May, the concentrations of lead, cobalt, chromium, cadmium, and nickel were 0.0656 mg/g, 0.0736 mg/g, 0.257 mg/g, 0.0532 mg/g, and 0.6782 mg/g respectively. In June, the concentration of lead decreased slightly to 0.0628 mg/g, while the concentrations of cobalt and cadmium also decreased to 0.0708 mg/g and 0.0494 mg/g respectively. The concentration of chromium increased to 0.3904 mg/g, while the concentration of nickel decreased to 0.613 mg/g. In July, there was a further decrease in the concentrations of all heavy metals, with lead, cobalt, chromium, cadmium, and nickel recording concentrations of 0.431 mg/g, 0.0515 mg/g, 0.25425 mg/g, 0.02675 mg/g, and 0.06 mg/g respectively.

Table 3: Monthly Concentrations (mg/g) of Heavy Metals in liver of Benthic fishes

	May	June	July
LEAD	0.0618	0.0618	0.864
COBALT	0.073	0.073	0.06175
CHROMIUM	0.348	0.348	0.4885
CADMIUM	0.0494	0.0494	0.0335
NICKEL	0.8102	0.8102	0.0665

Table 4: Monthly Concentrations (mg/g) of Heavy Metals in liver of Pelagic fishes

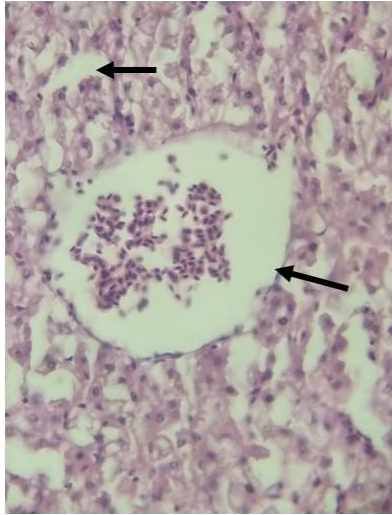
	May	June	July
LEAD	0.0656	0.0628	0.431
COBALT	0.0736	0.0708	0.0515
CHROMIUM	0.257	0.3904	0.25425
CADMIUM	0.0532	0.0494	0.02675
NICKEL	0.6782	0.613	0.06

3.4. Pathology of Liver of benthic and Pelagic fishes obtained from Ikpoba River

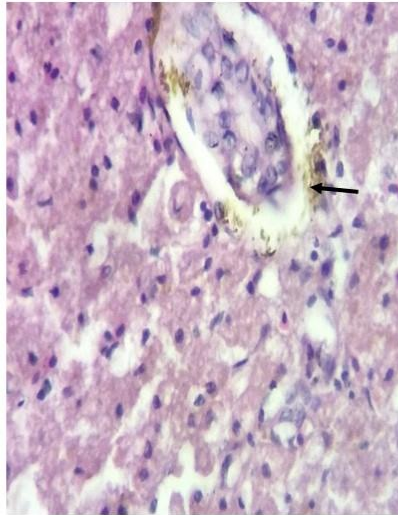
Histopathology of liver tissues of *Tilapia zilli* obtained from Ikpoba river showed some interesting results: In May, (Fig 2) liver tissues showed a congested central vein with mononuclear cells (long arrow). There are diffused mononuclear infiltrates (short arrow) visible around the central vein. The tissue appears to have a moderate level of cellular congestion, and the structure of hepatocytes is still identifiable. On the other hand, in June (Fig 2), the liver tissue exhibits a congested central vein (long arrow), similar to May but appears more pronounced. Diffused mononuclear infiltrates in the liver parenchyma (short arrow) are visible, indicating an inflammatory response. Marked steatosis is evident, with hepatocytes showing significant fat accumulation. Visible sinusoids can be observed amidst the inflamed tissue. In July (Fig 2), the central vein is not only congested but filled with inflammatory and mononuclear cells (long arrow), indicating an escalated inflammatory response over time. Hepatocytes reveal prominent steatosis (short arrow), suggesting progressive damage or

stress to the liver cells. The overall tissue structure appears more disorganized compared to previous months due to increased inflammation and steatosis.

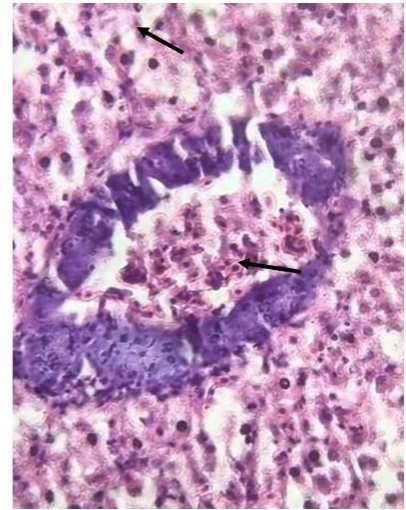
Typically, healthy liver tissues would exhibit clear, unobstructed central veins, absence of mononuclear infiltrate presence indicating no inflammation, and hepatocytes free of marked steatosis showcasing normal metabolic activity [40, 41]



Liver tissues of *Tilapia zilli* obtained from Ikpoba River in May



Liver tissues of *Tilapia zilli* obtained from Ikpoba River in June)



Liver tissues of *Tilapia zilli* obtained from Ikpoba in

Fig 2: Histopathology of liver tissues in *Tilapia zilli* obtained from Ikpoba river

3.5.Pathology of Liver d Benthic fishes obtained from Ikpoba River

In May, the liver tissue of *Carias Sp* shows (Fig 3) a congested central vein with mononuclear and inflammatory cells. This suggests an immune response, possibly due to an infection or other form of liver injury. There are also focal mononuclear infiltrates in the liver parenchyma, indicating localized areas of inflammation. The sinusoids are visible, which is normal, but the hepatocytes show marked steatosis. Steatosis refers to the abnormal retention of lipids within cells, suggesting a metabolic imbalance.

In June (Fig 3), the central vein remains congested, which could be due to increased blood flow or blockage. The liver parenchyma shows dispersed mononuclear infiltrates, suggesting a more widespread inflammatory response. The sinusoids and hepatocytes are visible, which is normal. However, there is focal pigmentation, which could indicate localized areas of cell damage or death. In July (Fig 3), the central vein is still congested, and there are visible mononuclear exudates, suggesting an ongoing immune response. The hepatocytes show prominent fatty changes, indicating metabolic disturbances, possibly due to conditions like non-alcoholic fatty liver disease (NAFLD) or alcoholic liver disease (ALD). There are also diffused mononuclear cells in the sinusoids, suggesting a widespread immune response.

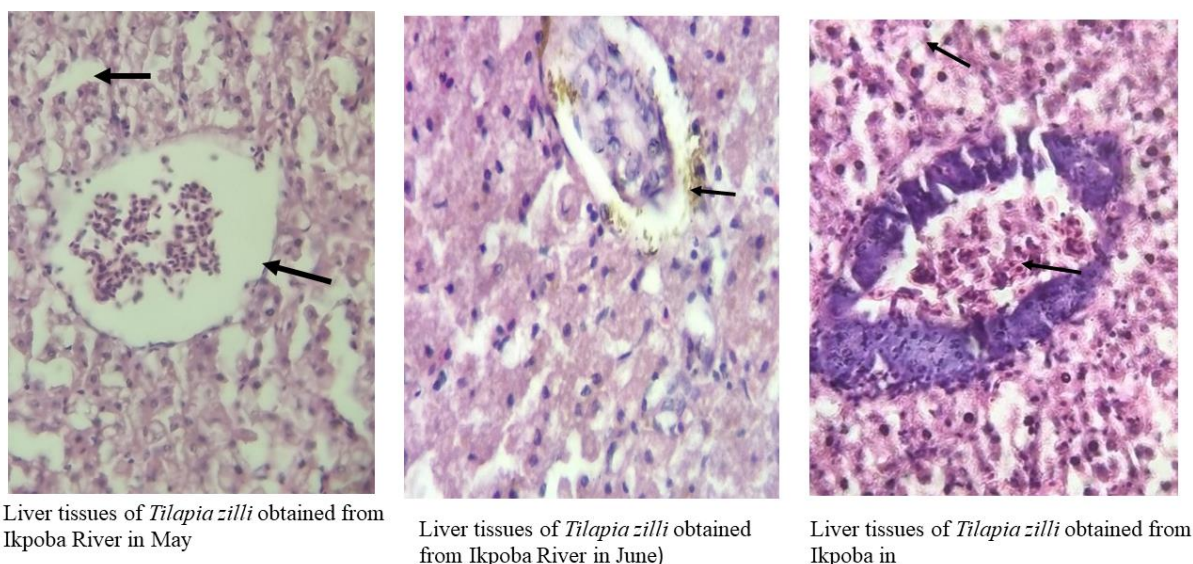


Fig 3: Histopathology of liver tissues in *Clarias sp* obtained from Ikpoba river

3.6. Discussion

3.6.1. Heavy metals in water samples

The data presented in Table 1 indicates the presence of heavy metals such as nickel, chromium, and cobalt in the Ikpoba River. The concentrations of these metals vary between upstream and downstream locations over the months of June, July, and August. Notably, the levels of lead and cadmium remained constant at 0 mg/L, suggesting that these metals may not be a significant concern in this context. However, the presence of chromium upstream is significant, which could have implications for the ecosystem and ecological health of the river [42].

Heavy metals in water bodies can pose serious threats to both aquatic life and human health [43, 44]. These metals, such as iron, copper, zinc, lead, cadmium, chromium, and mercury, can contaminate water sources through various sources like industrial effluents, mining waste, and agricultural runoff [45, 46]. These metals can cause a variety of issues in aquatic species, such as decreased hatching rates, physical abnormalities, and bioaccumulation in tissues [47, 48]. Moreover, they can also indirectly impact human health, as humans consume fish and other aquatic organisms that have accumulated these metals [43, 46].

Comparing these findings with other studies, it's evident that heavy metal contamination in water bodies is a global issue [46, 49-52]. For instance, a study conducted on the Pearl River in China found that heavy metals posed elevated ecological risks to certain aquatic organisms due to their presence in the water [53]. Another study by Kumar, et al. [50], highlighted the contamination of water with heavy metals in various water bodies throughout the world from 1994 to 2019. Similarly, Prasad, et al. [54] in another study conducted in the Garhwal Himalaya, India, found varying concentrations of heavy metals in potable water.

Looking specifically at previous studies conducted on the Ikpoba River, similar findings of heavy metal contamination have been reported. A study conducted in 2012 found that the concentrations of heavy metals were higher in effluents than in the water [55]. Another study conducted on the lower Ikpoba River reported that the concentrations of heavy metals in the water and sediment showed a seasonal pattern of variation [56]. Other studies including Tawari-Fufeyin [57], Igboanugo, et al. [58] and Wangboje and Ekundayo [59] have also reported varying concentrations

of heavy metals in waters samples from Ikpoba river. These studies, along with the current data, underscore the ongoing issue of heavy metal contamination in the Ikpoba River.

3.6.2. Heavy metals in sediment samples

The heavy metal concentrations in the sediments of the Ikpoba River have significant implications for the ecosystem and ecological health, particularly for aquatic organisms. Heavy metals, even in low quantities, can be harmful due to their toxicity, non-degradability, and high bioaccumulative potential [42, 60]. These metals can lead to various ailments in fish, such as reduced hatching rates and bioaccumulation in tissues [60]. The contamination of sediments with heavy metals can significantly impact the food chain, indirectly affecting human health when individuals consume contaminated fish [61].

The presence of heavy metals in river sediments worldwide has been reported in several studies, indicating that heavy metal pollution in aquatic environments is a global concern. These studies span across various countries. For instance, in Bangladesh, the presence of heavy metals was reported by Ali, et al. [62], Bhuyan, et al. [63], and Islam, et al. [64]. Similar findings were reported in China by Tang, et al. [53], Zheng, et al. [65] and Hoang, et al. [66] reported the same in Taiwan, while Jaskuła and Sojka [67] and Sojka and Jaskuła [61], did so in Poland. In Southern Africa, Ouma, et al. [68] reported these findings, as did Gupta, et al. [10] in India, Pesantes, et al. [69] in Ecuador, and Rezapour, et al. [70] in Iran. When comparing these findings with other studies, it becomes evident that heavy metal pollution in aquatic environments is indeed a global concern.

Specifically, on the Ikpoba River, there have been numerous studies conducted over the years that have reported on the heavy metal concentrations in its sediments. These studies have provided valuable insights into the environmental health of the river. For instance, the studies by Osa-Igwehide, et al. [28] (2016), Imiuwa, et al. [71], and Oguzie [56] have all contributed to our understanding of this issue. More recent studies by Ogbeide and Edene [72], as well as, Enuneku and Ineh [29], Enuneku and Ineh [73], have continued this important work, further emphasizing the need for ongoing monitoring and management of the river's health.

3.6.3. Heavy Metal Concentration in Fish Liver from Ikpoba River

The presence of heavy metals in the liver of both benthic and pelagic fishes from Ikpoba River is a clear indication of potential environmental contamination and possess a threat to humans who consume them [74-76]. The fluctuations in metal concentrations over the months in the liver of benthic and pelagic fishes from the Ikpoba River could be attributed to changes in water quality or variations in the fishes' diet and metabolism [77, 78]. These variations may reflect the dynamic nature of metal uptake and accumulation in fish tissues, influenced by environmental factors and biological processes [79]. Studies have shown that heavy metals can exhibit temporal variations in water bodies, affecting the bioavailability and accumulation of metals in aquatic organisms [80, 81]. The bioaccumulation of heavy metals in fish liver tissues can result from prolonged exposure to contaminated environments, leading to potential health risks for both fish and humans consuming these contaminated fish [82, 83].

Research on *Clarias gariepinus* and *Tilapia* fish across various global locations has revealed alarming concentrations of metals in fish tissues, exceeding the recommended limits for human consumption. This research spans multiple regions and rivers, each contributing unique insights into the issue.

In Nigeria, studies were conducted in the Elebele River [84], Ona River in Ibadan [85], Ogun River in Ogun State [86], New Calabar River [87], Asa River in Ilorin, Kwara State [88], Ose River in Ondo State [17] and Ikpoba river [28, 74].

In Egypt, the River Nile was studied [89], as well as other locations [90, 91]. Other studies were conducted in Lake Hayq, South Wollo, Ethiopia [92], and the Lake of Bhopal [93]. These findings indicate significant bioaccumulation of heavy metals in fish tissues, raising concerns about the potential health risks associated with consuming contaminated fish. [17, 74, 84].

When comparing the metal concentrations in benthic and pelagic fishes, it appears that benthic fishes tend to have higher concentrations of certain metals like lead and nickel, especially in July. This is consistent with findings by Odigie, et al. [74]. Benthic fishes, residing closer to the riverbed, tend to accumulate higher concentrations of certain metals like lead and nickel compared to pelagic fishes, which inhabit the water column [94]. This is attributed to the proximity of benthic fishes to the riverbed, where heavy metals often settle, leading to increased metal accumulation in their bodies [95]. The bioaccumulation of heavy metals in fish tissues, including lead and nickel, is influenced by their habitat and feeding behaviour [96]. In aquatic ecosystems, benthic fish species like large-scale tongue sole exhibit higher levels of metal accumulation in their edible tissues due to their benthic lifestyle [97]. Conversely, pelagic fishes may have lower exposure to these contaminants due to their habitat in the water column, potentially resulting in comparatively lower metal concentrations in their bodies [98].

The histopathology of the liver tissues of both tilapia and zebrafish shows signs of inflammation and steatosis, which are common responses to stress or injury, such as that caused by heavy metal exposure [99, 100]. The progression of these conditions over time indicates ongoing exposure to contaminants [101-103]. The histopathological findings in the liver tissues of *Tilapia zilli* and *Clarias species* from Ikpoba River indicate a progressive inflammatory response and steatosis, reflecting ongoing stressors impacting liver health [104]. These observations are crucial as fish liver health serves as a bioindicator for overall aquatic ecosystem health [104]. The liver, being a principal detoxifying organ, is highly susceptible to pollutants, as seen in fish exposed to crude oil pollution [105]. Additionally, dietary exposure to toxic metals like cadmium, chromium, lead, and mercury can lead to liver damage in fish, highlighting the impact of environmental contaminants on aquatic organisms [106]. Therefore, the observed liver pathologies in these fish species could signify broader ecological issues related to water quality and environmental stressors, emphasizing the importance of monitoring and addressing these concerns for ecosystem and ecological health.

3.6.4. Potential Sources of heavy metal in Ikpoba river

The Ikpoba River faces heavy metal pollution from various sources like industrial effluent discharge, waste management, urban runoff, and agricultural activities [28, 107]. Chromium presence in water, sediment, and fish liver samples indicates ecosystem-wide pollution, primarily from industrial effluent disposal, waste management, and urban runoff from Benin City [108]. Agricultural practices using heavy metal-containing fertilizers or pesticides also contribute to the contamination [107]². Effective regulation and mitigation of these sources are crucial for safeguarding the river's ecosystem and the communities dependent on it [28, 29, 109].

4.0. Conclusion

The research on the impact of urban runoff on fish fauna in the Ikpoba River has revealed significant findings, indicating potential environmental contamination from heavy metals in water, sediment, and fish liver samples. Fluctuations in metal concentrations in fish livers suggest changes in water quality or diet metabolism. Liver histopathology of tilapia and zebrafish shows signs of inflammation and steatosis, common responses to heavy metal exposure. These observations are crucial as fish liver health reflects overall aquatic ecosystem health. Recommendations include establishing a long-term monitoring program, identifying and regulating pollution sources, adopting an ecosystem-based approach, implementing policies, engaging stakeholders, conducting broader

pollution impact studies, and enhancing local capacity in environmental monitoring and management to protect the Ikpoba River's ecosystem and promote sustainable development.

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